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The Relationship between Xanthine Oxidoreductase and Xanthine Oxidase Activities in Plasma and Kidney Dysfunction

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Abstract

Background: Oxidative stress (OS) is thought to play a role in detrimental events among patients with chronic kidney disease (CKD). Although the mechanism of OS increases in CKD patients is unclear, it has been suggested that increased activity of xanthine oxidase (XO), the superoxide-producing form of xanthine oxidoreductase (XOR), plays a large role in enhancing OS. Therefore, we measured the activities of plasma XOR and XO among CKD patients.

Methods: Thirteen CKD patients were enrolled in this study. Their plasma XOR and XO activities were measured using a liquid chromatography-triple quadrupole mass spectroscopy (LC-TQMS) method, and the ratio of XO to XOR (XO/XOR) was calculated.

Results: The XOR and XO activities (pmol/h/mL plasma) were 8.30–58.2 and 9.15–40.8, and XO/XOR value was 0.698–1.240. Both XOR and XO showed positive correlation with estimated glomerular filtration rate (eGFR) (R=0.751, P=0.0020 and R=0.773, P=0.0031, respectively). On the other hand, XO/XOR showed negative correlation with eGFR (R=-0.614, P=0.0236).

Conclusion: XOR and XO activities are decreased, and XO/XOR ratio is increased, in accordance with renal dysfunction. High XO/XOR ratio in advanced CKD patients might contribute to the elevation of OS.

Keywords: Chronic kidney disease; Endothelium; Oxidative stress; Xanthine oxidase; Xanthine oxidoreductase

Introduction

Renal dysfunction is an independent risk factor for death and/or cardiovascular disease (CVD) events among the general population [1,2]. Although the reason why renal dysfunction increases CVD mortality and morbidity is unclear, the accumulated evidence suggests the major participation of oxidative stress (OS) [3]. Among patients with chronic kidney disease (CKD), OS increases in correlation with renal dysfunction [4], and enhanced OS among CKD patients induces an increase of CVD mortality and morbidity [5]. Such OS is widely considered as therapeutic target [6].

On the other hand, the participation of increased activity of xanthine oxidase (XO) on enhanced OS has been suggested [7]. XO is one of the isoforms of xanthine oxidoreductase (XOR, EC 1.2.3.2), which is known as a house-keeping enzyme. XOR is found as xanthine hydrogenase (XDH) in almost all living things, including bacteria, plants, eumycetes, and animals, while only mammals have both the XDH and XO forms [8]. XDH and XO are single-gene products [9] that catalyze the oxidation of hypoxanthine to xanthine and xanthine to uric acid; however, these two forms utilize different electron acceptors. Namely, XDH utilizes nicotinamide adenine dinucleotide (NAD⁺) and does not produce reactive oxygen species (ROS), whereas XO utilizes oxygen and generates ROS such as superoxide anion radicals and hydrogen peroxide. Accordingly, an increase in the XO/XOR ratio could accelerate OS via ROS generation.

Sakuma et al. reported that XDH → XO conversion is caused by disulfide compounds via reversible or non-reversible oxidation of thiol group in XDH [10]. Because thiol oxidation of serum albumin is enhanced in correlation with kidney dysfunction [4], plasma XO/XOR, which reflects XO/XOR of the capillary endothelium [11], might also be increased among advanced CKD patients. Moreover, no existing studies have

examined the relationship between plasma XOR or XO activity and renal function, for XOR activity is very low in healthy human and thus its precise measurement had been difficult [12,13].

The aim of this study is to evaluate the relationship between plasma XOR, XO and renal function.

Material and Methods

Thirteen predialysis CKD patients at Fukushima Medical University Hospital (Fukushima, Japan) were enrolled in the study. The patients' profiles including underlying kidney disease are shown in Table 1. No patient had received antioxidant agents such as ascorbic acid or vitamin E. Patients on corticosteroid therapy and those with malignancy were excluded from this study.

Physical measurements including height, weight, and blood pressure were taken, and then blood samples were collected to measure XOR and XO activities; 2 mL was drawn from the plasma obtained by centrifugation at 4°C immediately after the blood sampling from each patient, and stored at -80°C for 2–6 weeks until analysis. In addition, serum creatinine and C-reactive protein level, and plasma level of XOR substrates (hypoxanthine, xanthine and uric acid) were measured. As a marker of renal function, the estimated glomerular filtration rate (eGFR) was calculated by the CKD-EPI equation modified for Japanese using Japanese coefficient [14].

Measurement of XOR and XO activities was performed by the method previously reported by Murase et al. using liquid chromatography combined with triple quadrupole mass spectrometry (LC-TQMS method) [15]. Briefly, specimens were purified by Sephadex G25 column and were added to Tris buffer (pH 8.5) containing [¹³C₂,¹⁵N₂]xanthine, NAD⁺, and [¹³C₃,¹⁵N₃]uric acid. These mixtures were incubated at 37°C for 90 min. Subsequently, the mixtures were treated and the production levels of [¹³C₂,¹⁵N₂]uric acid were measured using liquid chromatography (NANOSPACE SI-2, Shiseido)-triple quadrupole mass spectrometer (TSQ-Quantum, Thermo Fisher Scientific) (LC-TQMS). XO activity was determined by measuring [¹³C₂,¹⁵N₂]uric acid formation in the absence of NAD⁺. XOR and XO activities were expressed as [¹³C₂,¹⁵N₂]uric acid in pmol/h/mL plasma. All measurements were duplicated and the mean values were adapted for the evaluation. Based on the XOR and XO activities obtained using the procedures described above, the ratio of XO to XOR (XO/XOR) was calculated.

Substrates of XOR (hypoxanthine, xanthine and uric acid) were measured using LC/MS method. Other biochemical tests were measured using standard laboratory techniques.

Values were expressed as means ± standard deviation unless otherwise stated. We used the statistical software Stat View 9.3 (SAS Institute Inc., Cary, NC, USA). To determine the magnitude of the correlation, we used Pearson's correlation coefficient (R). Item-category data (gender, primary disease, prescription, past history) were introduced into the analysis as dummy variables. The correlation was determined to be significant when the p-value was less than 0.05(5%) with Fisher's Z transformation.

The study was performed according to the principles of the Declaration of Helsinki and was approved by the local ethics committee (Fukushima Medical University, approval no. 2349). All subjects provided their written informed consent.

Table 1: Patient characteristics.

Age years	62.1 ± 18.8
Gender %male	53.8
BMI kg/m ²	24.2 ± 4.3
Primary CKD n	
Chronic glomerulonephritis	9
Nephrosclerosis	2
Polycystic kidney	1
Others	1
History of cardiovascular disease %	7.7
Antihypertensive medication %	84.6
Usage of erythrocyte-stimulating factor %	15.4
Usage of xanthine-oxidoreductase inhibitor %	61.5
a) None n	5
b) Febuxostat n	5
c) Topiroxostat n	1
d) Allopurinol n	2
Blood pressure mmHg	
Systolic	136 ± 22
Diastolic	84 ± 6
Estimated glomerular filtration rate mL/min/1.73 m ²	42.7 ± 23.4
C-reactive protein	0.14 ± 0.17

Results

Table 2 shows the results of measurement regarding XOR and XO activities, and these substrates. The XOR and XO activities (pmol/h/mL plasma) were 8.30–58.2 and 9.15–40.8, and XO/XOR value was 0.698–1.240.

The relationship between XOR, XO, XO/XOR and renal function (eGFR) is shown in Figure 1. A significantly positive correlation between eGFR and both XOR and XO was determined (R=0.751, P=0.0020 and R=0.773, P=0.0031, respectively). On the other hand, XO/XOR showed significantly negative correlation with eGFR (R=-0.614, P=0.0236), indicating that decreased renal function contributes to XDH → XO conversion.

Table 3 shows the results of simple regression analysis, and Table 4 shows that of multiple (forward stepwise) regression analysis between XOR, XO, XO/XOR and clinical data, in which clinical data were determined to be the explanatory variables, and XOR, XO and XO/XOR were the criterion variables. In the multiple regression analysis regarding XOR, eGFR and

prescription of XOR inhibitor were adopted as an independent significant explanatory variable. Same variables (eGFR and prescription of XOR inhibitor) were also adopted independently in the multiple regression analysis regarding XO. These results suggest that low eGFR and usage of XOR inhibitor suppresses both XOR and XO activities. As to the multiple regression analysis regarding XO/XOR, selected explanatory variables were eGFR and xanthine.

Table 2: The results of measurement regarding xanthine oxidoreductase (XOR) and xanthine oxidase (XO) activities, and these substrates.

	Mean	SD	Range
XOR activity pmol/h/mL plasma	23.1	15.9	8.30-58.2
XO activity pmol/h/mL plasma	19.4	10.0	9.15-40.8
XO/XOR	0.940	0.183	0.698-1.24
Hypoxanthine μ M	2.27	2.02	0.30-7.09
Xanthine μ M	5.60	7.57	0.11-26.4
Uric acid μ M	220.8	74.4	23.5-350.0

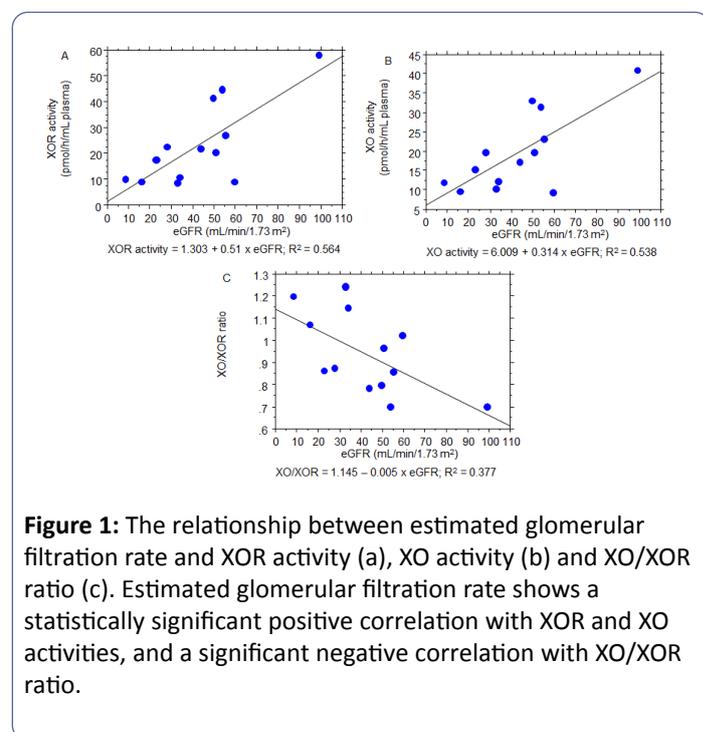
Table 3: The simple regression between XOR, XO, XO/XOR and clinical data, * $P < 0.05$.

	XOR		XO		XO/XOR	
	R	P-value	R	P-value	R	P-value
Estimated glomerular filtration rate	0.751	0.0020*	0.733	0.0031*	-0.614	0.0236*
Hypoxanthine	0.003	0.9912	-0.060	0.8505	-0.105	0.7381
Xanthine	-0.613	0.0239*	-0.637	0.0171*	0.712	0.0048*
Uric acid	-0.562	0.0443*	-0.555	0.0479*	0.346	0.2533
Age	-0.288	0.3492	-0.279	0.3644	0.246	0.4261
Gender (male=1)	-0.375	0.2131	-0.396	0.1855	0.418	0.1597
BMI	0.005	0.9864	0.003	0.9931	-0.092	0.7714
History of cardiovascular disease	0.665	0.0113*	0.640	0.0166*	-0.393	0.1886
Antihypertensive medication	-0.545	0.0530	-0.554	0.0485*	0.391	0.1919
Usage of erythrocyte-stimulating factor	-0.260	0.4008	-0.260	0.4010	0.220	0.4788
Usage of xanthine-oxidoreductase inhibitor	-0.740	0.0027*	-0.770	0.0013*	0.597	0.0296*
Systolic blood pressure	0.096	0.7613	0.144	0.6471	-0.050	0.8730

Diastolic blood pressure	-0.262	0.3955	-0.195	0.5328	0.220	0.4789
C-reactive protein	0.109	0.7303	0.164	0.6006	0.009	0.9766

Table 4: The multiple (stepwise) regression between XOR, XO, XO/XOR and clinical data, * $P < 0.05$.

	XOR		XO		XO/XOR	
	R	P-value	R	P-value	R	P-value
R of the test	0.866	0.0010*	0.874	0.0007*	0.815	0.0043*
Estimated glomerular filtration rate	0.514	0.0171*	0.472	0.0226*	-0.422	0.0556
Xanthine	-----	-----	-----	-----	0.569	0.0152*
Uric acid	-----	-----	-----	-----	-----	-----
History of cardiovascular disease	-----	-----	-----	-----	-----	-----
Antihypertensive medication	-----	-----	-----	-----	-----	-----
Usage of xanthine-oxidoreductase inhibitor	-0.493	0.0210*	-0.542	0.0114*	-----	-----



Discussion

In this report, we demonstrated that both XOR and XO activities are decreased, and $XDH \rightarrow XO$ conversion of plasma XOR is accelerated, in accordance with renal dysfunction. To the best of our knowledge, this is the first report to reveal the

relationship between renal function and XOR activity including conformation change, i.e. XDH \rightarrow XO conversion.

XOR is a house-keeping enzyme and is recognized as a urate-synthesizer in a clinical medicine context. In the present study, we measured plasma XOR. The XOR state of plasma is thought to reflect that of capillary endothelial cells, since XOR is abundantly found in capillary endothelium and scarce in the endothelium of major vessels and endocardium [16]. The total area of the endothelium in humans is equal to that of 6 tennis courts, with most of the endothelium covering capillary vessels [17]. Capillary XOR plays a major role in redox maintenance of the whole body [16,18] and takes part in various acute pathophysiologies, such as ischemia-reperfusion injury via XDH \rightarrow XO conversion [9,19,20]. Besides, XDH \rightarrow XO conversion is essential for the lactation process, a fundamental function of mammals [21].

In this study, we demonstrated clear relationship between eGFR and both XOR and XO activity. Otaki et al. reported that both high and low XOR activity (≥ 120 and <33 pmol/100 μ L/h), measured using a combination of [$^{13}C_2$, $^{15}N_2$] xanthine and liquid chromatography/triple quadrupole mass spectrometry, was related to poor outcome in patients with chronic heart disease [22]. Although the pathological significance by which abnormal XOR activity is unclear, too low XOR activity might reflect impaired endothelial viability.

XOR inhibitor has potential to reduce detrimental event among CKD patients [7,23], and the conserving function of the XOR inhibitor on endothelial cells is independent of the urate-lowering effect [24]. About 85% of XOR exists as XDH in reductive condition such as cytosol of hepatic cell [25].

Not like XDH, XO utilizes molecular oxygen as electron acceptor and generates ROSs such as superoxide, which causes tissue injury including endothelial dysfunction. We have hypothesized that the ratio of XO over XOR increases in accordance with renal dysfunction in oxidative condition such as plasma. In this context, the progress of CKD among subjects with hyperuricemia [26-28] might be explained by the profound elevation of XO/XOR ratio.

In the multiple regression analysis regarding XO/XOR ratio, plasma xanthine level was selected as dependent explanatory variable. Although this result is interesting, the meaning of it is uncertain at the present.

In the present study, XO/XOR ratio exceeded 1 in patients with severely-impaired renal function. We suppose that this phenomenon reflects the irreversible modification of XOR, which could not be recovered by NAD⁺ treatment. XOR exhibits XDH \rightarrow XO conversion not only by reversible disulfide bridge formation, but also by irreversible modification caused by proteases [29,30]. Such "irreversible" XO modification might cause this spurious elevation of XO/XOR ratio to over 1.

In this study, we revealed a "decrease" of both XOR and XO activities in accordance with renal dysfunction. This result is in line with the report by Otaki et al., in which eGFR of "low XOR" group (53 ± 25 mL/min/1.73 m²) was significantly lower than that of "normal XOR" group (68 ± 26 mL/min/1.73 m²) [21]. On

the other hand, Gondouin et al. reported the "increase" of XOR activity in accordance with renal dysfunction [31]. To measure XOR activity, Gondouin et al. used the "classical method", in which XOR activity is measured as urate level after administration of xanthine to the specimen. This classical method might well overestimate XOR activity because of elevated endogenous urate and other substances.

This study has several limitations. The first limitation is its cross-sectional nature: a prospective study is needed to prove the effect of the XOR inhibitor on XO/XOR ratio. The second is that the degree of endothelial function or vascular damage is not evaluated in this study. The third and perhaps most obvious limitation of this report is that the number of participants was small (13 participants). Studies with a larger subject population are warranted to allow more detail statistical analysis including comparison test between stratified groups and following post-hoc test.

In conclusion, we demonstrated that XOR and XO activities are decreased, and XDH \rightarrow XO conversion of plasma XOR is accelerated, in accordance with renal dysfunction. Clinical strategies targeting XO/XDH modification should be encouraged in the future, for high XO/XOR ratio in advanced CKD patients might contribute to the elevation of OS.

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References

1. Go AS, Chertow GM, Fan D, McCulloch CE, Hsu CY (2004) Chronic kidney disease and the risks of death, cardiovascular events, and hospitalization. *N Engl J Med* 351: 1296-1305.
2. Nakayama M, Metoki H, Terawaki H, Ohkubo T, Kikuya M, et al. (2007) Kidney dysfunction as a risk factor for first symptomatic stroke events in a general Japanese population--the Ohasama study. *Nephrol Dial Transplant* 22: 1910-1915.
3. Himmelfarb J, Stenvinkel P, Ikizler TA, Hakim RM (2002) The elephant in uremia: oxidant stress as a unifying concept of cardiovascular disease in uremia. *Kidney Int* 62: 1524-1538.
4. Terawaki H, Yoshimura K, Hasegawa T, Matsuyama Y, Negawa T, et al. (2004) Oxidative stress is enhanced in correlation with renal dysfunction: examination with the redox state of albumin. *Kidney Int* 66: 1988-1993.
5. Terawaki H, Takada Y, Era S, Funakoshi Y, Nakayama K, et al. (2010) The redox state of albumin and serious cardiovascular incidence in hemodialysis patients. *Ther Apher Dial* 14: 465-471.
6. Ciccone MM, Cortese F, Gesualdo M, Carbonara S, Zito A, et al. (2013) Dietary intake of carotenoids and their antioxidant and anti-inflammatory effects in cardiovascular care. *Mediators Inflamm* 2013: 782137.
7. Terawaki H, Nakayama M, Miyazawa E, Murata Y, Nakayama K, et al. (2013) Effect of allopurinol on cardiovascular incidence among hypertensive nephropathy patients: the Gonryo study. *Clin Exp Nephrol* 17: 549-553.

8. Nishino T (1994) The conversion of xanthine dehydrogenase to xanthine oxidase and the role of the enzyme in reperfusion injury. *J Biochem* 116: 1-6.
9. Hille R, Nishino T (1995) Flavoprotein structure and mechanism. 4. Xanthine oxidase and xanthine dehydrogenase. *FASEB J* 9: 995-1003.
10. Sakuma S, Fujita J, Nakanishi M, Wada SI, Fujimoto Y (2008) Disulfide S-monoxides convert xanthine dehydrogenase into oxidase in rat liver cytosol more potently than their respective disulfides. *Biol Pharm Bull* 31: 1013-1016.
11. Houston M, Estevez A, Chumley P, Aslan M, Marklund, et al. (1999) Binding of xanthine oxidase to vascular endothelium. *J Biol Chem* 19: 4985-4994.
12. Newaz MA, Adeeb NN (1998) Detection of xanthine oxidase in human plasma. *Med J Malaysia* 53: 70-75.
13. Murase T, Nampei M, Oka M, Ashizawa N, Matsumoto K, et al. (2016) Xanthine oxidoreductase activity assay in tissues using stable isotope-labeled substrate and liquid chromatography high-resolution mass spectrometry. *J Chromatogr B Analyt Technol Biomed Life Sci* 1008: 189-197.
14. Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF, et al. (2009) A new equation to estimate glomerular filtration rate. *Ann Intern Med* 150: 604-612.
15. Murase T, Nampei M, Oka M, Miyachi A, Nakamura T (2016) A highly sensitive assay of human plasma xanthine oxidoreductase activity using stable isotope-labeled xanthine and LC/TQMS. *J Chromatograph. B* 1039: 51-58.
16. Jarasch ED, Grund C, Bruder G, Heid HW, Keenan TW, et al. (1981) Localization of xanthine oxidase in mammary-gland epithelium and capillary endothelium. *Cell* 25: 67-82.
17. Higashi Y, Noma K, Yoshizumi M, Kihara Y (2009) Endothelial function and oxidative stress in cardiovascular diseases. *Circ J* 73: 411-418.
18. Matsuyama Y, Hayashi T, Terawaki H, Negawa T, Terada T, et al. (2009) Human astrocytes and aortic endothelial cells actively convert the oxidized form of albumin to the reduced form: reduced albumin might participate in redox regulation of nerve and blood vessel systems. *J Physiol Sci.* 59: 207-215.
19. McCord JM (1985) Oxygen-derived free radicals in postischemic tissue injury. *N Engl J Med* 312: 159-163.
20. Saugstad OD (1988) Hypoxanthine as an indicator of hypoxia: its role in health and disease through free radical production. *Pediatr Res* 23: 143-150.
21. Vorbach C, Scriven A, Capecchi MR (2002) The housekeeping gene xanthine oxidoreductase is necessary for milk fat droplet enveloping and secretion: gene sharing in the lactating mammary gland. *Genes Dev* 16: 3223-3235.
22. Otaki Y, Watanabe T, Kinoshita D, Yokohama M, Takahashi T, et al. (2017) Association of plasma xanthine oxidoreductase activity with severity and clinical outcome in patients with chronic heart failure. *Int J Cardiol* 228: 151-157.
23. Goicoechea M, de Vinuesa SG, Verdalles U, Ruiz-Caro C, Ampuero J, et al. (2010) Effect of allopurinol in chronic kidney disease progression and cardiovascular risk. *Clin J Am Soc Nephrol* 5: 1388-1393.
24. George J, Carr E, Davies J, Belch JJ, Struthers A (2006) High-dose allopurinol improves endothelial function by profoundly reducing vascular oxidative stress and not by lowering uric acid. *Circulation* 114: 2508-2516.
25. Sakuma S, Fujita J, Nakanishi M, Wada S, Fujimoto Y (2008) Disulfide s-monoxides convert xanthine dehydrogenase into oxidase in rat liver cytosol more potently than their respective disulfides. *Biol Pharm Bull* 31: 1013-1016.
26. Iseki K, Ikemiya Y, Inoue T, Iseki C, Kinjo K, et al. (2004) Significance of hyperuricemia as a risk factor for developing ESRD in a screened cohort. *Am J Kidney Dis* 44: 642-650.
27. Obermayr RP, Temml C, Gutjahr G, Knechtelsdorfer M, Oberbauer R, et al. (2008) Elevated uric acid increases the risk for kidney disease. *J Am Soc Nephrol* 19: 2407-2413.
28. Kawashima M, Wada K, Ohta H, Terawaki H, Aizawa Y (2011) Association between asymptomatic hyperuricemia and new-onset chronic kidney disease in Japanese male workers: a long-term retrospective cohort study. *BMC Nephrol* 12: 31.
29. Chung HY, Baek BS, Song SH, Kim MS, Huh JJ, et al. (1997) Xanthine dehydrogenase/xanthine oxidase and oxidative stress. *Age* 20: 127-140.
30. Sarnesto A, Linder N, Raivio KO (1996) Organ distribution and molecular forms of human xanthine dehydrogenase/xanthine oxidase protein. *Lab Invest.* 74: 48-56.
31. Gondouin B, Jourde-Chiche N, Sallee M, Dou L, Cerini C, et al. (2015) Plasma xanthine oxidase activity is predictive of cardiovascular disease in patients with chronic kidney disease, independently of uric acid levels. *Nephron* 131: 167-174.